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2 **Isolation and characterization of tetrachloroethylene and**  
3 ***cis*-1,2-dichloroethylene-dechlorinating propionibacteria**

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13  
14 **Running Title: Isolation and characterization of propionibacteria for dechlorination**

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Abstract

Two rapidly growing propionibacteria that could reductively dechlorinate tetrachloroethylene (PCE) and *cis*-1,2-dichloroethylene (*cis*-DCE) to ethylene were isolated from environmental sediments. Metabolic characterization and partial sequence analysis of their 16S rRNA genes showed that the new isolates, designated as strains *Propionibacterium* sp. HK-1 and *Propionibacterium* sp. HK-3, did not match any known PCE or *cis*-DCE-degrading bacteria. Both strains dechlorinated relatively high concentrations of PCE (0.3 mM) and *cis*-DCE (0.52 mM) under anaerobic conditions without accumulating toxic intermediates during incubation. Cell-free extracts of both strains catalyzed PCE and *cis*-DCE dechlorination; degradation was accelerated by the addition of various electron donors. PCE dehalogenase from strain HK-1 was mediated by a corrinoid protein since the dehalogenase was inactivated by propyl iodide only after reduction by titanium citrate. The amounts of chloride ions (0.094 mM and 0.103 mM) released after PCE (0.026 mM) and *cis*-DCE (0.05 mM) dehalogenation using the cell-free enzyme extracts of both strains, HK-1 and HK-3, were stoichiometrically similar (91% and 100%), indicating that PCE and *cis*-DCE were fully dechlorinated. Radiotracer studies with [1, 2-<sup>14</sup>C] PCE and [1, 2-<sup>14</sup>C] *cis*-DCE indicated that ethylene was the terminal product: partial conversion to ethylene was observed. Various chlorinated aliphatic compounds (PCE, trichloroethylene, *cis*-DCE, *trans*-1, 2-dichloroethylene, 1, 1-dichloroethylene, 1, 1-dichloroethane, 1, 2-dichloroethane, 1, 2-dichloropropane, 1, 1, 2-trichloroethane, and vinyl chloride) were degraded by cell-free extracts of strain HK-1.

**Key words:** Tetrachloroethylene, *cis*-1,2-dichloroethylene, chlorinated aliphatic compounds, corrinoid, Propionibacteria, PCE dehalogenase

1  
2 **Introduction**

3  
4 Tetrachloroethylene (PCE), an effective and widely used degreasing solvent and fumigant, is a  
5 frequently detected recalcitrant xenobiotic pollutant in soil and groundwater around the world [19]. PCE  
6 can be reductively dechlorinated by anaerobic microorganisms to trichloroethylene (TCE) and  
7 *cis*-1,2-dichloroethylene (*cis*-DCE) [1, 7, 16, 26, 38, 45, 48, 51]. However, for anaerobic bioremediation  
8 to be useful, PCE must be degraded to nonchlorinated, environmentally harmless products.  
9 Environmental accumulation of *cis*-DCE and vinyl chloride (VC) is undesirable, because *cis*-DCE is a  
10 suspected carcinogen and VC is toxic. Some bacteria have been isolated, including *Dehalococcoides*  
11 *ethenogenes* strain 195 and *Dehalococcoides* sp. BAV1, FL2, and GT, that can dechlorinate *cis*-DCE via  
12 VC to ethane [21, 22, 37, 46, 48]. The strain 195 is the only known microorganism capable of  
13 dechlorinating PCE to ethene. Similarly, mixed cultures capable of complete conversion of PCE or TCE  
14 to ethene invariably contain organisms closely related to *Dehalococcoides*. Hendrickson *et al.* [24]  
15 demonstrated the importance of *Dehalococcoides* in bioremediation by conducting an extensive survey  
16 of the presence of *Dehalococcoides* at multiple contaminated sites. The collected evidence from many  
17 studies has led to such great interest in the *Dehalococcoides* group of microorganisms.

18 Hydrocarbon biodegradation can be carried out by a variety microorganisms; activity is generally  
19 thought to be ubiquitous. This was also supposed for the biodegradation of PCE and TCE to *cis*-DCE.  
20 However, the surprising finding from more than a decade of research is that only one species of bacteria  
21 within the genus *Dehalococcoides* has been isolated that can dechlorinate PCE and *cis*-DCE to ethene.  
22 However, it would be imprudent to assume that no organisms outside the *Dehalococcoides* group may be  
23 capable of the reductive dehalogenation of PCE and *cis*-DCE to innocuous compounds such as ethane.

24 There is, therefore, interest in the discovery and identification of anaerobes capable of converting PCE  
25 and *cis*-DCE without the accumulation of harmful by-products. There is also interest in characterizing the  
26 degradation reactions. This work reports the dehalogenation of PCE and *cis*-DCE by cell-free extracts of  
27 two isolated microorganisms (*Propionibacterium* sp. HK-1 and *Propionibacterium acnes* HK-3). Data are  
28 shown that, for the first time, indicate bacterial degradation of PCE by Propionibacteria may be possible  
29 without the accumulation of toxic intermediates.

30  
31  
32 **Materials and Methods**

33  
34 **Chemicals**

35  
36 All chemicals were of analytical grade and purchased from Kanto Chemical (Tokyo, Japan). *cis*-DCE  
37 was from Tokyo Chemical Industry (Tokyo, Japan) and other chlorinated chemicals (PCE, TCE,  
38 *trans*-1,2-dichloroethylene (*trans*-DCE), 1,1-dichloroethylene (1,1-DCE), 1,1-dichloroethane,  
39 1,2-dichloroethane, 1,2-dichloropropane, 1,1,2-trichloroethane, and VC) were purchased from GL

1 science (Tokyo, Japan). [1, 2-<sup>14</sup>C] PCE and [1, 2-<sup>14</sup>C] *cis*-DCE were purchased from Sigma Chemical  
2 Co. (St. Louis, Mo., USA).

### 3 4 Enrichment and Isolation

5  
6 40 soil samples were collected aseptically in Hokkaido, Japan from near a factory of electrical parts,  
7 near a dry-cleaner's and from around a livestock farm. 5 sewage sludge samples were also taken.  
8 Organisms in the samples were enriched and cultivated under anaerobic conditions by inoculating 3 g  
9 samples into 50-ml serum bottles with 30 ml MY medium of the following composition (per liter of  
10 deionized water): K<sub>2</sub>HPO<sub>4</sub>, 0.2 g; NaCl, 0.05 g; MgSO<sub>4</sub>·7H<sub>2</sub>O, 0.2 g; CaCl<sub>2</sub>, 0.05 g; FeCl<sub>3</sub>·6H<sub>2</sub>O,  
11 0.0083 g; MnCl<sub>2</sub>·4H<sub>2</sub>O, 0.014 g; NaMoO<sub>4</sub>·2H<sub>2</sub>O, 0.017 g; ZnCl<sub>2</sub>, 0.001 g; yeast extract, 2.0 g; glucose,  
12 2.0 g; *L*-cystein, 0.5 g; resazurin, 0.001 g; at pH 7.2. After autoclaving 30 ml of the medium, the  
13 headspaces of the bottles were purged with N<sub>2</sub> gas (>99.9%) for 15 min, and sealed with Teflon-lined  
14 rubber septa and aluminium crimp caps. PCE and *cis*-DCE were injected into the bottles via  
15 microsyringe at final concentrations of 0.06 mM and 0.1 mM, respectively. Samples were incubated at  
16 30 °C and 120 rpm for 30 days in the dark. To identify cultures that did not accumulate toxic  
17 by-products of PCE and *cis*-DCE, the headspaces of the sample bottles were periodically analyzed by  
18 gas chromatography. Only two enrichment cultures, from ditch sludge of a dry cleaning shop (Muran,  
19 Hokkaido, Japan), did not accumulate toxic by-products of PCE and *cis*-DCE from all the samples tested.  
20 Whenever PCE and *cis*-DCE were degraded, cultures were used to inoculate second-generation cultures,  
21 which were then used subsequently to inoculate third-generation cultures, and so on, up to six successive  
22 generations. To isolate colonies, 10-fold dilution of the enrichment culture was spread on petri plates  
23 containing MY medium with 1.5% agar. A sterile glass tube (0.4 mm i.d×20 mm), filled with cotton  
24 fiber soaked with PCE (0.06 mM) or *cis*-DCE (0.1 mM) solution was attached to the inside lid of the  
25 petri plate. The plate was then incubated under anaerobic conditions using Anaerobic Gas Generation  
26 Kit (Oxoid Ltd, Hants, UK). The procedure was repeated twice to ensure pure cultures. By the  
27 above-mentioned isolation procedure, two representative PCE and *cis*-DCE-degrading bacteria were  
28 successfully isolated.

### 29 30 DNA sequencing and phylogenetic analysis

31  
32 For the phylogenetic identification of the two isolates, 16S rRNA gene fragments were amplified by  
33 polymerase chain reaction (PCR) with a pair of universal primers, 27f and 1392r under standard  
34 conditions. The PCR mixture contained 1 μL 10 pmol each primer, 5 μL 10×Ex Taq buffer, 4 μL 2.5  
35 mM each dNTP, 0.25 μL Takara Ex Taq HS (TAKARA BIO, Shiga, Japan), 2 μL DNA extract in a final  
36 volume of 50 μL. After initial denaturation at 94 °C for 3 min, 30 cycles of denaturation at 94 °C for 30  
37 s were performed. Primer annealing at 55 °C for 1 min and extension at 72 °C for 1 min followed. A  
38 final extension was then carried out at 72 °C for 7 min. The PCR product was purified with an  
39 ExoSAP-IT (GE Healthcare) PCR purification kit and sequenced using BigDye Terminator v1.1 Cycle  
40 Sequencing kit (Applied Biosystems, Foster City, CA, USA) and an ABI PRISM 3130 Genetic Analyzer

1 (Applied Biosystems, Foster City, CA, USA). The sequences determined in this study were compared  
2 with other gene sequences in the NCBI database using the BLAST program  
3 (<http://www.ncbi.nlm.nih.gov/BLAST/>). The sequences determined in this study and data retrieved from  
4 the GeneBank database were aligned using CLUSTAL W. The alignments were refined by visual  
5 inspection. A neighbor-joining tree was constructed using a TreeView software package. Bootstrap  
6 analysis was used to estimate the reliability of phylogenetic reconstructions (1000 replicates).  
7 Physiological characteristics of the isolates were also determined by commercially available  
8 identification systems (API 20 A; bioMérieux, Japan).

#### 9 10 Degradation experiments

11  
12 The kinetics of degradation of PCE (0.06, 0.12, 0.3, and 0.6 mM) and *cis*-DCE (0.1, 0.21, 0.52, and 0.98  
13 mM) were determined at 30 °C using 20 ml MY medium in 50 mL serum bottles, covered with  
14 Teflon-lined caps. Two isolated strains were pre-grown on MY medium for 72 h. Pre-cultures were  
15 harvested by centrifugation (8000×g, 4 °C, 10 min), washed twice with 50 mM phosphate buffer (pH  
16 7.2), and suspended in MY medium. 200 µL cell suspension inoculated in the medium had an optical  
17 density at 590 nm (OD<sub>590</sub>) of 0.03. The gas phase was N<sub>2</sub>. Samples were incubated at 30 °C and 120 rpm  
18 for two weeks.

#### 19 20 Preparation of cell extracts

21  
22 Two isolates were pre-grown on MY medium for 96 h. Pre-cultures were harvested by centrifugation  
23 (8000×g, 4 °C, 10 min), and suspended in MY medium. 2 ml cell suspension (0.33 mg protein/ ml) was  
24 transferred to a 120 mL serum bottle containing 78 mL fresh medium. The culture medium and headspace  
25 of the bottle were aseptically purged with N<sub>2</sub> for 15 min. Samples were incubated at 30 °C and 120 rpm  
26 for 72 h. Cells (1.2 g wet weight) were harvested by centrifugation (8000×g, 4 °C, 15 min), resuspended  
27 in 6 ml 50 mM Tris-HCl (pH 7.5) containing 2 mM dithiothreitol (DTT), and, 5% glycerol, and lysed in  
28 an ice bath, using a BRANSON (Danbury, CT, U.S.A.) ultrasonic disrupter SONIFIER 250 at 30 s flash  
29 for 5 min. Unbroken cells and debris were recovered by centrifugation (18800×g) and resonicated. Both  
30 supernatants were pooled and filtered through a 0.22 µm filter (ADVENTEC, DISMIC-25AS, Bedford,  
31 USA). The filtrate served as the enzyme extract.

#### 32 33 Enzyme assays

34  
35 Enzyme activity was determined through PCE and *cis*-DCE degradation. Assays were carried out in 20  
36 mL serum bottles equipped with Teflon septa. The assay mixture comprised: 1 mL enzyme, 3 mL 50 mM  
37 Tris-HCl buffer (pH 7.5), 2 mM DTT and 5 mM glucose. The headspace was purged with N<sub>2</sub> gas. PCE  
38 (0.12 mM) or *cis*-DCE (0.21 mM) were then injected and incubated at 30 °C for a maximum of 2 h. The  
39 reaction was terminated by adding 0.2 mL 5 M H<sub>2</sub>SO<sub>4</sub>. Headspace samples were analyzed by gas

1 chromatography. Protein was quantified by the Bradford method using the Bio-Rad protein assay reagent  
2 (Bio-Rad, Hercules, California, U.S.A.).

#### 3 4 Effect of electron donors

5  
6 The effects of electron donors on PCE and *cis*-DCE degradation was studied. 1 mL enzyme (0.79 mg  
7 protein mL<sup>-1</sup>) was transferred to a 20 ml serum bottle containing 3 ml 50 mM Tris-HCl (pH 7.5) and DTT  
8 (2 mM). The influences of various potential electron donors (ethanol, methanol, glucose, yeast extract,  
9 pyruvate, fumarate, acetate, lactate, methyl viologen and formate) were examined at final concentrations  
10 of 5 mM; except for yeast extract, which was used at 2.0 g/L). Hydrogen (0.9 μM) was tested at a partial  
11 pressure of  $0.5 \times 10^5$  Pa. The bottles were purged with N<sub>2</sub> gas and sealed with Teflon-coated stoppers  
12 before PCE (0.12 mM) and *cis*-DCE (0.21 mM) were added.

#### 13 14 Degradation of other aliphatic compounds

15  
16 Enzymes were used at 1.2 mg protein mL<sup>-1</sup> and compared with controls, containing no enzyme. The initial  
17 concentration of each compound was 0.12 mM. After 1 hour, the degradations of PCE, TCE, *cis*-DCE,  
18 *trans*-DCE, 1,1-DCE, 1,1-dichloroethane, 1,2-dichloroethane, 1,2-dichloropropane, 1,1,2-trichloroethane,  
19 and VC were determined by gas chromatography.

#### 20 21 Effect of propyl iodide on PCE degradation

22  
23 Cell extract (1 mL; 1.2 mg protein mL<sup>-1</sup>) was added to 3 ml 0.1 M Tris-HCl (pH 7.5) in a glass bottle  
24 wrapped in aluminium foil. Dehalogenase activity was measured in the test system described above  
25 except that titanium (III) citrate (2 mM) was used instead of DTT. Propyl iodide, 1-iodopropane (PI; 0.5  
26 mM) was added and the cell extract was illuminated (250 W lamp) for 5 min.

#### 27 28 Gas chromatographic analysis of substrates

29  
30 PCE and TCE were identified and quantified by static-headspace analysis using a gas chromatograph.  
31 PCE, TCE and DCE isomers in 250 μL headspace samples were determined using a GC-8A gas  
32 chromatograph (Shimadzu Co., Japan) equipped with a flame ionization detector (FID) and a glass  
33 column (i.d. 3.2 φ×2.1 m; Silicone DC-550 20% Chromosorb W [AWDMCS] 80/100). The column was  
34 maintained at 60 °C. The injector and detector were kept at 140 °C. The gas samples were analyzed two  
35 to three times to verify reproducibility. Ethylene, ethane, and carbon dioxide were analyzed by gas  
36 chromatography using a Shimadzu GC-14B, equipped with a glass column, Unibeads C 60/80 (i.d. 3.2  
37 φ×2.1 m), and a thermal conductivity detector (TCD). Helium was used as a carrier gas. GCMS analysis  
38 was conducted with a Shimadzu GC/MS system (GCMS-QP2010) and an Rxi-5 ms capillary column  
39 (30 m, 0.25 mm ID, 1.00 μm *df*; Restek, Pennsylvania, USA). The column temperature program during  
40 GC-MS analysis of metabolites was as follows: held at 60 °C for 2 min, increased to 300 °C at 20 °C per



1 min, and held at 300 °C for 5 min. The injection, interface, and ion-source temperatures were 280, 280,  
2 and 250 °C, respectively. Helium (99.995%) was used as a carrier gas at a flow rate of 1.0 ml min<sup>-1</sup>.

### 3 4 Degradation of [1, 2-<sup>14</sup>C] PCE and [1, 2-<sup>14</sup>C] *cis*-DCE

5 Microcosm preparation, sampling procedures, and methods for verification of <sup>14</sup>C-labeled volatile  
6 compounds (PCE, TCE, *cis*-DCE, VC, ethylene, and ethane) and <sup>14</sup>CO<sub>2</sub> are well described by Freedman  
7 and Gossett [14]. Degradations of [1, 2-<sup>14</sup>C] PCE and [1, 2-<sup>14</sup>C] *cis*-DCE by two isolates were performed  
8 in 120 ml serum bottles containing 50 ml of MY medium. Pre-cultures were inoculated in the medium.  
9 Thereafter, culture medium and the bottles' headspaces were purged with N<sub>2</sub> for 20 min and then spiked  
10 with 40,000 dpm [1, 2-<sup>14</sup>C] PCE (specific activity = 0.6 mCi mmol<sup>-1</sup>) or [1, 2-<sup>14</sup>C] *cis*-DCE (specific  
11 activity = 0.6 mCi mmol<sup>-1</sup>). Labelled PCE and *cis*-DCE were added along with the final addition of  
12 unlabeled PCE and *cis*-DCE. <sup>14</sup>C-labeled volatile compounds were analyzed with by GC-combustion  
13 [14]. After incubation for 10 days, the reaction was stopped by bringing the pH to 2.5 with perchloric  
14 acid (3M). Chlorinated compounds released by acidification were fractionated by GC and trapped  
15 individually in ScientiVerse-E liquid scintillation cocktail (Fisher Scientific). The CO<sub>2</sub> generated was  
16 absorbed by CO<sub>2</sub> absorption liquid (Carbo Sorb E, Perkin Elmer, USA). Radioactivity was measured in  
17 an LS 6500 liquid scintillation counter (Beckman Instruments, Inc., Fullerton, CA, USA). The total  
18 disintegrations per minute (dpm) in a bottle of each compound were calculated using the appropriate  
19 Henry constants [18].

### 20 21 Other analysis

22  
23 The concentration of chloride ions released during PCE and *cis*-DCE degradation was measured by an  
24 ionic chromatograph Dionex ICS-1000 equipped with a conductivity detector (Dionex Co., CA, USA),  
25 using a 4 mm anionic exchanger column, IonPack AS9-HC. The volume of injection was 25 µL. The  
26 mobile phase was 9 mM sodium carbonate solution with flow rate of 1 ml min<sup>-1</sup>.

### 27 28 Data analysis

29  
30 All results are indicated as mean values with standard deviations (±95% confidence interval) of triplicate  
31 experiments, except the experiment of PI effect on PCE degradation. Significant difference was  
32 determined by Student's *t* testing with *p* < 0.05.

### 33 34 Nucleotide sequence accession number

35  
36 The 16S rRNA gene sequences of the isolates (strain HK-1 and strain HK-3) determined in this study  
37 were deposited in the DDBJ under accession nos. AB540663 and AB540664, respectively.

## 38 39 **Results**

### 40 Taxonomy of isolated organisms

1  
2 The isolated strains were named HK-1 and HK-3. The PCE and *cis*-DCE-degrading organisms were  
3 anaerobic, Gram-positive, and rod-shaped bacteria. Colonies of each strain appeared white when  
4 cultured on MY agar plates with vapors of PCE (0.12 mM) or *cis*-DCE (0.21 mM). The strains showed  
5 similar physiological characteristics, they were not able to produce indole, and produced  $\beta$ -glucosidase  
6 but not protease (Table 1). The strains displayed positive catalase and oxidase activities but did not  
7 produce urease. Based on these characteristics, both strains were considered to belong to the genus  
8 *Propionibacterium*. As shown in the phylogenetic tree (Fig. 1), both strains were identified as  
9 *Propionibacterium* sp. 16S rRNA gene sequence analysis showed that strain HK-1 was closely related to  
10 *Propionibacterium acidipropionici* DH42 AY360222 (98.2% sequence identity). The 16S rRNA gene of  
11 the strain HK-3 completely agreed with that of *Propionibacterium acnes* W1392 AY642051 (100%  
12 sequence identity).

#### 13 14 Biodegradation of PCE and *cis*-DCE

15  
16 Figure 2 shows time courses of the ratios of residual to initial PCE and *cis*-DCE concentrations, and the  
17 cell densities (OD<sub>590</sub>) of liquid cultures of strains HK-1 and HK-3. Tests of PCE degradation by strain  
18 HK-1 at high concentrations (0.06, 0.12, 0.3, and 0.6 mM) showed that PCE was not degraded at 0.6  
19 mM. However, when its initial concentration was 0.3 mM, PCE degradation commenced after 24 h and  
20 was degraded up to 64% during the incubation period. Similar was observed at 0.06 and 0.12 mM, but  
21 PCE degradation was slower than at 0.3 mM PCE. The amount of biomass increased during cultivation.  
22 *cis*-DCE was not degraded at 0.98 mM by HK-3, though it was rapidly degraded at all other tested  
23 concentrations (0.1, 0.21, and 0.52 mM). PCE and *cis*-DCE were not the sole carbon/energy sources for  
24 the growth of these strains (data not shown).

#### 25 26 Effect of electron donors and chloride release

27  
28 The rate of PCE degradation varied with different electron donors (Fig. 3). Methyl viologen, glucose,  
29 and yeast extract enhanced PCE degradation. Other electron donors (hydrogen, lactate, and acetate) were  
30 slightly more specific than the control for PCE degradation. Although methyl viologen most effectively  
31 enhanced PCE degradation, it was not used in subsequent experiments because of concerns for human  
32 safety. Similar results were obtained on *cis*-DCE degradation experiment. Based on these results,  
33 glucose was chosen as the most effective electron donor for PCE and *cis*-DCE degradation. To  
34 determine whether chloride ions were stoichiometrically released during PCE and *cis*-DCE degradation,  
35 the chloride ion contents in the enzyme extracts of both strains were determined (Fig. 4). Chloride  
36 concentrations generated from PCE after 20, 40 and 60 minutes were 0.022, 0.057, and 0.093,  
37 respectively, indicating that *ca.* 91% of stoichiometric chloride ions were generated from PCE (Fig. 4  
38 (A)). Therefore, most of the PCE was dechlorinated by the cell-free extracts of strain HK-1. Chloride  
39 concentrations from *cis*-DCE after 20, 40, and 60 minutes were 0.019, 0.084, and 0.096, respectively,  
40 consistent with the complete dechlorination of *cis*-DCE (Fig. 4 (B)).

1  
2 Inactivation of PCE degradation by propyl iodide

3  
4 The effects of propyl iodide and titanium citrate on PCE degradation by the cell-free enzyme extract are  
5 presented in Fig. 5. The extract lost 80% of its initial PCE degradation activity upon incubation with  
6 propyl iodide and titanium (III) citrate in the dark within 10 min. Subsequent exposure to light restored  
7 80% of the activity within 20 min. Titanium citrate in the absence of propyl iodide did not have any  
8 inhibitory effect and no inhibition was recorded with propyl iodide alone (data not shown).

9  
10 Degradation of other aliphatic compounds

11  
12 The degradation of other halogenated aliphatic compounds by crude enzyme was investigated. PCE  
13 pregrown enzyme resulted in degradations of PCE, TCE, *cis*-DCE, *trans*-DCE, 1,1-DCE,  
14 1,1-dichloroethane, 1,2-dichloroethane, 1,2-dichloropropane, 1,1,2-trichloroethane, and VC of 20%,  
15 16% , 25%, 16%, 23%, 23%, 27%, 10%, 25%, and 30%, respectively (Table 2). When *cis*-DCE  
16 pregrown enzyme was tested, the degradations of *cis*-DCE, *trans*-DCE, 1, 1-DCE, and VC were as 35%,  
17 18%, 25%, and 40%, respectively. However, PCE, TCE, 1, 1-dichloroethane, 1, 2-dichloroethane, 1,  
18 2-dichloropropane, and 1, 1, 2-trichloroethane showed no degradation after extended incubation (6 h).  
19 The degradation of VC was more than that of PCE or *cis*-DCE in reactions with PCE and *cis*-DCE crude  
20 enzyme.

21  
22 Degradation of [1, 2-<sup>14</sup>C] PCE and [1, 2-<sup>14</sup>C] *cis*-DCE

23  
24 Radioisotope experiments were performed with [1, 2-<sup>14</sup>C] PCE. Radioactivities in 10-day cultures were  
25 recovered in ethylene (7%), and cells (8%) in the radioisotope experiments. Although only 15% of  
26 radioactivity was recovered in ethylene, the mechanism was concluded to involve reductive  
27 dechlorination since the intermediate (ethylene) of reductive dechlorination was observed. Radioisotope  
28 experiments were also performed with [1, 2-<sup>14</sup>C] *cis*-DCE, where radioactivity was only recovered in  
29 ethylene (4%). Other metabolites (TCE, *cis*-DCE, VC, and ethane) were not detected. Negligible  
30 amounts of radioactivity (<1%) was observed in cells.

31  
32 **Discussion**

33 PCE and *cis*-DCE-degrading anaerobic bacteria, *Propionibacterium* sp. HK-1 and *Propionibacterium* sp.  
34 HK-3 were isolated and characterized. 16S rRNA gene sequencing revealed that strains HK-1 and HK-3  
35 were not similar to any of the following known PCE or *cis*-DCE-degrading bacteria: *Desulfuromonas*  
36 *michiganensis* BB1 (AF357915) [48], *Desulfuromonas michiganensis* BRS1 (AF357915) [48],  
37 *Dehalococcoides* sp. H10 (AY914178) [50], *Desulfitobacterium* sp. B31e3 (AB289347) [56],  
38 *Geobacter lovleyi* SZ (AY914177) [49], *Geobacter lovleyi* GT (AY914178) [[50], *Sulfurospirillum*  
39 *halorespirans* PCE-M2 (AF218076) [35], *Desulfitobacterium hafniense* Y51 (AB049340) [51],  
40 *Desulfitobacterium hafniense* JH1 (EU523374) [13], *Dehalococcoides ethenogens* 195 (AF004928) [37],

1 *Desulfitobacterium* sp. PCE-S (AJ512772) [39], *Desulfitobacterium* sp. PCE1 (X81032) [16],  
2 *Desulfitobacterium* sp. KBC1 (AB194704) [54], *Desulfitobacterium hafniense* TCE1 (X95742) [17],  
3 *Dehalobacter restrictus* (U84497) [27], *Sulfurospirillum multivorans* (X82931) [45], *Desulfuromonas*  
4 *chloroethenica* (U49748) [31], *Desulfitobacterium dehalogenans* (L28946) [55], *Desulfomicrobium*  
5 *norvegicum* (AJ277897) [25], *Gammaproteobacterium* MS-1 (L43508) [47], *Dehalococcoides* sp. FL2  
6 (AF357918) [23], *Dehalococcoides* sp. BAV1 (AY165308) [22], *Desulfitobacterium* sp. Viet-1  
7 (AF357919) [33], *Desulfitobacterium hafniense* PCP-1 (U40078) [10], *Desulfitobacterium*  
8 *chlororespirans* (U68528) [34, 44], *Desulfomonile tiedjei* DCB-1 (M26635) [40], *Desulfitobacterium*  
9 *hafniense* DCB-2 (AY013365) [32], *Clostridium* sp. KYT-1 (AB214911) [30], *Acetobacterium woodii*  
10 (DD223101) [52], *Rhodococcus* sp. Sm-1 (DQ834672) [42], *Xanthobacter flavus* (DQ834674) [28].  
11 Although several electron donors effectively enhanced PCE and *cis*-DCE dechlorination, glucose and  
12 yeast extract were the most effective. No growth or reductive dechlorination of PCE or *cis*-DCE was  
13 observed when glucose or yeast extract were eliminated from the MY medium. This observation suggests  
14 that glucose and yeast extract served sources of nutrients and electrons.

15 Reductive dechlorination of PCE has been achieved by many pure cultures belonging to four different  
16 metabolic groups: halorespirers, acetogens, methanogens, and facultative anaerobes [9]. To date, only  
17 *Dehalococcoides ethenogenes* strain 195 has been known to degrade PCE to nontoxic ethene, while  
18 most other natural biotic processes degrade PCE to toxic products [1, 7, 16, 26, 37, 38, 45, 48, 51, 56].  
19 PCE was long thought to be non-biodegradable in the presence of oxygen, but PCE degradation by  
20 *Pseudomonas stutzeri* OX1, involving a toluene-o-xylene monooxygenase has been reported [43].  
21 Aerobic PCE degradation has also been reported by the white-rot fungus *Trametes versicolor* [11].  
22 Reductive dehalogenation has been shown to be a major mechanism in the complete degradation of PCE,  
23 though other reactions or mechanisms may also facilitate the complete degradation of PCE. An  
24 anaerobic oxidative degradation of *cis*-DCE and VC under Fe(III)-reducing and humic acid-reducing  
25 conditions has been reported [3, 4, 5]. Under such conditions, no dechlorinated ethenes were detected  
26 during the experiments [3, 4, 5].

27 In this study, no chlorinated ethenes were observed during PCE degradation with cells and cell-free  
28 extracts of strain HK-1. *Ca.* 91% of the stoichiometric amount of chloride ions was generated during the  
29 incubation of cell-free extracts containing PCE. This indicates that most of the PCE was dechlorinated.  
30 Strain HK-3 released chloride ions stoichiometrically with *cis*-DCE dehalogenation, indicating that  
31 *cis*-DCE was fully dechlorinated. Similar results were observed in *Clostridium* species. Hata *et al.* [20]  
32 and Kim *et al.* [30] reported that *Clostridium* species, such as *Clostridium* sp. DC1, *Clostridium*  
33 *butyricum* NBRC 3315, *Clostridium acetobutylicum* NBRC 13949, and *Clostridium* sp. KYT-1, also  
34 showed ability to degrade *cis*-DCE without any accumulation of VC or ethylene. The two isolates were  
35 able to dechlorinate PCE and *cis*-DCE in the presence and absence of Fe(III) in the MY medium,  
36 respectively. This finding supports that the anaerobic degradation of chlorinated ethenes is not linked to  
37 the reduction of Fe (III) to Fe (II) [20]. A similar result was demonstrated by *Clostridium* sp. DC1,  
38 which has also been reported to dechlorinate *cis*-DCE in the presence or absence of Fe (III) [20]. The  
39 origin of the carbon in the CO<sub>2</sub> gas, whether from PCE or from the materials in the medium, was studied.  
40 Radiotracer studies with [1, 2-<sup>14</sup>C] PCE and [1, 2-<sup>14</sup>C] *cis*-DCE showed that ethylene was the terminal

1 product; partial conversion to ethylene was observed. However, the unrecovered reaction products of  
2 PCE and *cis*-DCE remain unknown. This requires further study to elucidate the mechanism and  
3 products.

4 Anaerobic halorespiring bacteria (dehalorespirers) are widely accepted as being important to biologic  
5 dechlorination in anoxic environments. Compared with dehalorespirers, co-metabolic PCE  
6 dechlorination proceeds more slowly. The rates of PCE dechlorination by *Methanosarcina* sp. and  
7 *Acetobacterium woodii* were  $3.5 \times 10^{-5}$  and  $3.6 \text{ nmol h}^{-1} \cdot \text{mg protein}^{-1}$ , respectively [12]. In comparison,  
8 PCE dechlorination by *Sulfurospirillum multivorans*, *Dehalococcoides ethenogenes* strain 195,  
9 *Clostridium bifermentans* DPH-1, and *Dehalobacter restrictus* (strain PER-K23) has been reported to  
10 occur at 3.0, 4.14, 0.4, and  $1.0 \mu\text{mol h}^{-1} \cdot \text{mg protein}^{-1}$  [7, 26, 45]. Dechlorination by strain HK-1 occurred  
11 at a rate ( $18 \text{ nmol h}^{-1} \cdot \text{mg protein}^{-1}$ ) close to that of bacteria which degrade PCE co-metabolically but  
12 lower than that of dehalorespires. However, the PCE dechlorination rate was higher than that of aerobic  
13 white-rot fungus *Trametes versicolor* ( $2.75 \text{ nmol h}^{-1} \cdot \text{mg protein}^{-1}$ ) [11]. In Figure 2, at the highest  
14 concentrations of PCE and *cis*-DCE, there was no degradation but substantial growth was observed (cell  
15 growth with high PCE), possibly evidence of a co-metabolic process.

16 Protein-bound cobalamin from a number of anaerobic organisms e.g., *Desulfitobacterium hafniense*  
17 Y51, *Dehalococcoides ethenogenes*, *Desulfitobacterium* sp. strain PCE-1, *Clostridium bifermentans*  
18 DPH-1, *Desulfitobacterium* sp. PCE-S, *Sulfurospirillum multivorans*, *Sporomusa ovata*  
19 (homoacetogenic strain), *Dehalobacter restrictus*, and *Methanosarcina thermophila* have been reported  
20 to degrade PCE [3, 8, 29, 41 52]. It has been postulated that enzyme-bound cobalamin is in a  
21 superreduced state [cob(I)alamin], in which the alkyl residue of an alkyl halide can bind to the cobalt  
22 atom [15]. The same mechanism has been applied to the binding of the propyl chain of propyl iodide to  
23 cobalt, thus inactivating the enzyme [6]. This work clearly shows that the mechanism of PCE  
24 degradation by PCE dehalogenase from *Propionibacterium* sp. HK-1 is mediated by a corrinoid protein,  
25 since the dehalogenase was inactivated by propyl iodide only after reduction by titanium citrate.

26 The capability to degrade halogenated aliphatic compounds has mainly been observed in the genera  
27 *Desulfitobacterium*, *Clostridium*, and the related *Dehalobacter restrictus* [2, 8, 27, 30, 38, 41]. This work  
28 reports the first demonstration of a broad spectrum of chlorinated aliphatics being degraded by cell-free  
29 extracts of *Propionibacterium* sp. HK-1. Biodegradation of xenobiotic pollutants by *Propionibacterium*  
30 sp. is not known well. However, a *Propionibacterium* capable of degrading *O*-aryl alkyl ethers and  
31 various aromatic hydrocarbons has been reported recently [53].

32 Two PCE and *cis*-DCE-degrading bacteria (strains HK-1 and HK-3) were isolated and characterized.  
33 Neither accumulated toxic intermediate compounds such as TCE, *cis*-DCE, or VC. This is the first  
34 report of anaerobic organisms capable of degrading PCE without the accumulation of chlorinated  
35 ethenes and with partially mineralization to ethylene. This may be useful for the development of  
36 biological remediation of chlorinated ethene-contaminated sites. Furthermore, the ability of strain HK-1  
37 to degrade several halogenated aliphatic compounds has potential to aid the amelioration of  
38 environments contaminated with mixtures of halogenated substances. Given that *Propionibacterium* sp.  
39 is a ubiquitous microorganism worldwide, it is an interesting model for further studies of PCE  
40 degradation.

1  
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6  
7 **References**

- 8 1. Adrian L, Szewzyk U, Wecke J, Gorisch H (2000) Bacterial dehalorespiration with chlorinated  
9 benzenes. *Nature* 408:580-583
- 10 2. Bouwer EJ, McCarty PL (1983) Transformation of 1- and 2-carbon halogenated aliphatic organic  
11 compounds under methanogenic conditions. *Appl Environ Microbiol* 45:1286-1294
- 12 3. Bradley PM, Chapelle FH (1997) Kinetics of DCE and VC mineralization under methanogenic and  
13 Fe(III)-reducing conditions. *Environ Sci Technol* 31:2692–2696
- 14 4. Bradley PM, Chapelle FH (1998) Microbial mineralization of VC and DCE under different terminal  
15 electron accepting conditions. *Anaerobe* 4:81–87
- 16 5. Bradley PM, Chapelle FH, Lovley DR (1998) Humic acids as electron acceptors for anaerobic  
17 microbial oxidation of vinyl chloride and dichloroethene. *Appl Environ Microbiol* 64:3102–3105
- 18 6. Brot N, Weissbach H (1965) Enzymatic synthesis of methionine; chemical alkylation of the  
19 enzyme-bound cobamide. *J Biol Chem* 240:3064-3070
- 20 7. Chang YC, Hatsu M, Jung K, Yoo YS, Takamizawa K (2000) Isolation and characterization of a  
21 tetrachloroethylene dechlorinating bacterium, *Clostridium bifermentans* DPH-1. *J Biosci Bioeng*  
22 89:489-491
- 23 8. Chang YC, Okeke BC, Hatsu M, Takamizawa K (2001) In vitro dehalogenation of  
24 tetrachloroethylene (PCE) by cell-free extracts of *Clostridium bifermentans* DPH-1. *Bioresource*  
25 *Technology* 78:141-147
- 26 9. Damborsky J (1999) Tetrachloroethene-dehalogenating bacteria. *Folia Microbiol* 44:247-262
- 27 10. Dennie D, Gladu I, Lepine F, Villemur R, Bisailon JG, Beaudet R (1998) Spectrum of the reductive  
28 dehalogenation activity of *Desulfitobacterium frapperi* PCP-1. *Appl Environ Microbiol*  
29 64:4603-4606
- 30 11. Ernest MU, Gabarrell X, Sarrà M, Caminal G, Vicent T, Reddy CA (2006) Novel aerobic  
31 perchloroethylene degradation by the white-rot fungus *Trametes versicolor*. *Environ Sci Technol*  
32 40:7796-7802
- 33 12. Fetzner S (1998) Bacterial dehalogenation. *Appl Microbiol Biotechnol* 50:633-657
- 34 13. Fletcher KE, Ritalahti KM, Pennell KD, Takamizawa K, Löffler FE (2008) Resolution of culture  
35 *Clostridium bifermentans* DPH-1 into two populations, a *Clostridium* sp. and  
36 tetrachloroethene-dechlorinating *Desulfitobacterium hafniense* strain JH1. *Appl Environ Microbiol*  
37 74:6141-6143
- 38 14. Freedman DL, Gossett JM (1989) Biological reductive dechlorination of tetrachloroethylene and  
39 trichloroethylene to ethylene under methanogenic conditions. *Appl Environ Microbiol* 55:2144-2151
- 40 15. Friedrich W (1975) Vitamin B<sub>12</sub> und verwandte corrinoid. Thieme, Stuttgart.

- 1 16. Gerritse J, Renard V, Pedro Gomes TM, Lawson PA, Collins MD, Gottschal JC (1996)  
2 *Desulfitobacterium* sp. strain PCE1, an anaerobic bacterium that can grow by reductive dechlorination  
3 of tetrachloroethylene or *ortho*-chlorinated phenols. Arch Microbiol 165:132-140
- 4 17. Gerritse J, Drzyzga O, Kloetstra G, Keijmel M, Wiersum LP, Hutson R, Collins MD, Gottschal JC  
5 (1999) Influence of different electron donors and acceptors on dehalorespiration of tetrachloroethene  
6 by *Desulfitobacterium frappieri* TCE1. Appl Environ Microbiol 65:5212-5221
- 7 18. Gossett JM (1987) Measurement of Henry's law constants for C<sub>1</sub> and C<sub>2</sub> chlorinated hydrocarbons.  
8 Environ Sci Technol 21:202-208
- 9 19. Gossett JM (2002) Fishing for microbes. Science 298:974-975
- 10 20. Hata J, Miyata N, Kim ES, Takamizawa K, Iwahori K (2004) Anaerobic degradation of  
11 *cis*-1,2-dichloroethylene and vinyl chloride by *Clostridium* sp. Strain DC1 isolated from landfill  
12 leachate sediment. J Biosci Bioeng 97:196-201
- 13 21. He J, Ritalahti KM, Aiello MR, Löffler FE (2003) Complete detoxification of vinyl chloride by an  
14 anaerobic enrichment culture and identification of the reductively dechlorinating population as a  
15 *Dehalococcoides* species. Appl Environ Microbiol 69:996-1003
- 16 22. He J, Ritalahti KM, Yang KL, Koenigsberg SS, Löffler FE (2003) Detoxification of vinyl chloride to  
17 ethene coupled to growth of an anaerobic bacterium. Nature 424:62-65
- 18 23. He J, Sung Y, Krajmalnik-Brown R, Ritalahti KM, Löffler FE (2005) Isolation and characterization  
19 of *Dehalococcoides* sp. strain FL2, a trichloroethene (TCE)- and 1,2-dichloroethene-respiring  
20 anaerobe. Environ Microbiol 7:1442-1450
- 21 24. Hendrickson ER, Payne JA, Young RM, Starr MG, Perry MP, Fahnestock S, Ellis DE, Ebersole RC  
22 (2002) Molecular analysis of *Dehalococcoides* 16S ribosomal DNA from chloroethene-contaminated  
23 sites throughout north America and Europe. Appl Environ Microbiol 68:485-495
- 24 25. Hippe H, Vainshtein M, Gogotova GI, Stackebrandt E (2003) Reclassification of *Desulfobacterium*  
25 *macestii* as *Desulfomicrobium macestii* comb. Nov. Int J Syst Evol Microbiol 53:1127-1130
- 26 26. Holliger C, Schraa G, Stams AJM, Zehnder AJB (1993) A highly purified enrichment culture  
27 couples the reductive dechlorination of tetrachloroethene to growth. Appl Environ Microbiol  
28 59:2991-2997
- 29 27. Holliger C, Hahn D, Harmsen H, Ludwig W, Schumacher W, Tindall B, Vazquez F, Weiss N,  
30 Zehnder AJB (1998) *Dehalobacter restrictus* gen. nov. and sp. nov., a strictly anaerobic bacterium  
31 that reductively dechlorinates tetrachloroethene and trichloroethene in an anaerobic respiration. Arch  
32 Microbiol 169:313-321
- 33 28. Ito Y, Suzuki K, Ando T, Nashimoto K, Niwa K, Shimada K (2007) Technology for detection of  
34 degradation microorganisms and bioremediation. Matsushita Technical Journal 53:16-21
- 35 29. Jablonski PE, Ferry JG (1992) Reductive dechlorination of trichloroethylene by the CO-reduced CO  
36 dehydrogenase enzyme complex from *Methanosarcina thermophila*. FEMS Microbiol Lett 96:55-60
- 37 30. Kim ES, Nomura I, Hasegawa Y, Takamizawa K (2006) Characterization of a newly isolated  
38 *cis*-1,2-dichloroethylene and aliphatic compound-degrading bacterium, *Clostridium* sp. strain KYT-1.  
39 Biotechnol Bioprocess Eng 11:553-556

- 1 31. Krumholz LR (1997) *Desulfuromonas chloroethenica* sp. nov. uses tetrachloroethylene and  
2 trichloroethylene as electron acceptors. *Int J Syst Bacteriol* 47:1262-1263
- 3 32. Leadbetter JR (2005) *Environmental Microbiology*, Vol. 397, In: Löffler FE, Sanford RA, Ritalahti  
4 KM (eds), *Methods in Enzymology*, Elsevier-Academic Press, pp 77-111
- 5 33. Löffler FE, Tiedje JM, Sanford RA (1999) Fraction of electrons consumed in electron acceptor  
6 reduction and hydrogen thresholds as indicators of halorespiratory physiology. *Appl Environ*  
7 *Microbiol* 65:4049-4056
- 8 34. Löffler FE, Sanford RA, Tiedje JM (1996) Initial characterization of a reductive dehalogenase from  
9 *Desulfitobacterium chlororespirans* Co23. *Appl Environ Microbiol* 62:3809-3813
- 10 35. Luijten MLGC, Weert JD, Smidt H, Boschker HTS, Vos WMD, Schraa G, Stams AJM (2003)  
11 Description of *Sulfurospirillum halorespirans* sp. nov., an anaerobic, tetrachloroethene-respiring  
12 bacterium, and transfer of *Dehalospirillum multivorans* to the genus *Sulfurospirillum* as  
13 *Sulfurospirillum multivorans* comb. nov. *Int J Syst Evol Microbiol* 53:787-793
- 14 3. Maillard J, Schumacher W, Vazquez F, Regeard C, Hagen WR, Holliger C (2003) Characterization of  
15 the corrinoid iron-sulfur protein tetrachloroethene reductive dehalogenase of *Dehalobacter restrictus*.  
16 *Appl Environ Microbiol* 69:4628-4638
- 17 37. Maymó-Gatell X, Tandoi V, Gossett JM, Zinder SH (1997) Isolation of a bacterium that reductively  
18 dechlorinates tetrachloroethene to ethane. *Science* 276:1568-1571
- 19 38. Miller E, Wohlfarth G, Diekert G (1996) Studies on tetrachloroethene respiration in *Dehalospirillum*  
20 *multivorans*. *Arch Microbiol* 166:379-387
- 21 39. Miller E, Wohlfarth G, Diekert G (1998) Comparative studies on tetrachloroethene reductive  
22 dechlorination mediated by *Desulfitobacterium* sp. strain PCE-S. *Arch Microbiol* 168:513-519
- 23 40. Mohn WW, Kennedy KJ (1992) Reductive dehalogenation of chlorophenols by *Desulfomonile*  
24 *tiedjei* DCB-1. *Appl Environ Microbiol* 58:1367-1370
- 25 41. Neumann A, Wohlfarth G, Diekert G (1995) Properties of tetrachloroethene dehalogenase of  
26 *Dehalospirillum multivorans*. *Arch Microbiol* 163:276-281
- 27 42. Phelps TJ, Malachowsky K, Schram RM, White DC (1991) Aerobic mineralization of vinyl chloride  
28 by a bacterium of the order Actinomycetales. *Appl Environ Microbiol* 57:1252-1254
- 29 43. Ryoo D, Shim H, Canada K, Barbieri P, Wood TK (2000) Aerobic degradation of  
30 tetrachloroethylene by toluene-o-xylene monooxygenase of *Pseudomonas stutzeri* OX1. *Nat*  
31 *Biotechnol* 18:775-778
- 32 44. Sanford RA, Cole JR, Löffler FE, Tiedje JM (1996) Characterization of *Desulfitobacterium*  
33 *chlororespirans* sp. nov., which grows by coupling the oxidation of lactate to the reductive  
34 dechlorination of 3-chloro-4-hydroxybenzoate. *Appl Environ Microbiol* 62:3800-3808
- 35 45. Scholz-Muramatsu H, Neumann A, Meßmer M, Moore E, Diekert G (1995) Isolation and  
36 characterization of *Dehalospirillum multivorans* gen. nov., a tetrachloroethene-utilizing, strictly  
37 anaerobic bacterium. *Arch Microbiol* 163:48-56
- 38 46. Seshadri R, Adrian L, Fouts DE, Eisen JA, Phillippy AM, Methe BA, Ward NL, Nelson WC, Deboy  
39 RT, Khouri HM, Kolonay JF, Dodson RJ, Daugherty SC, Brinkac LM, Sullivan SA, Madupu R,  
40 Nelson KE, Kang KH, Impraim M, Tran K, Robinson JM, Forberger HA, Fraser CM, Zinder SH,



- 1 Heidelberg JF (2005) Genome sequence of the PCE-dechlorinating bacterium *Dehalococcoides*  
2 *ethenogenes*. Science 307:105-108
- 3 47. Sharma PK, McCarty PL (1996) Isolation and characterization of a facultatively aerobic bacterium  
4 that reductively dehalogenates tetrachloroethene to *cis*-1,2-dichloroethene. Appl Environ Microbiol  
5 62:761-765
- 6 48. Sung Y, Ritalahti KM, Sanford RA, Urbance JW, Flynn SJ, Tiedje JM, Löffler FE (2003)  
7 Characterization of two tetrachloroethene (PCE)-reducing, acetate-oxidizing anaerobic bacteria, and  
8 their description as *Desulfuromonas michiganensis* sp. nov. Appl Environ Microbiol 69:2964-2974
- 9 49. Sung Y, Fletcher KE, Ritalahti KM, Apkarian RP, Ramos-Hernández N, Sanford RA, Mesbah NM,  
10 Löffler FE (2006a) *Geobacter lovleyi* sp. nov. strain SZ, a novel metal-reducing and  
11 tetrachloroethene-dechlorinating bacterium. Appl Environ Microbiol 72:2775-2782
- 12 50. Sung Y, Ritalahti KM, Apkarian RP, Löffler FE (2006b) Quantitative PCR confirms purity of strain  
13 GT, a novel trichloroethene-to-ethene-respiring *Dehalococcoides* isolate. Appl Environ Microbiol  
14 72:1980-1987
- 15 51. Suyama A, Iwakiri R, Kai K, Tokunaga T, Sera N, Furukawa K (2001) Isolation and characterization  
16 of *Desulfitobacterium* sp. strain Y51 capable of efficient dechlorination of tetrachloroethene and  
17 polychloroethanes. Biosci Biotechnol Biochem 65:1474-1481
- 18 52. Terzenbach DP, Blaut M (1994) Transformation of tetrachloroethylene to trichloroethylene by  
19 homoacetogenic bacteria. FEMS Microbiol Lett 123:213-218
- 20 53. Thorenoor N, Kim YH, Lee CJ, Yu MH, Engesser KH (2009) A previously uncultured, paper mill  
21 *Propionibacterium* is able to degrade O-aryl alkyl ethers and various aromatic hydrocarbons.  
22 Chemosphere 75:1287-1293
- 23 54. Tsukagoshi N, Ezaki S, Uenaka T, Suzuki N, Kurane R (2006) Isolation and transcriptional analysis  
24 of novel tetrachloroethene reductive dehalogenase gene from *Desulfitobacterium* sp. strain KBC1.  
25 Appl Microbiol Biotechnol 69:543-553
- 26 55. Utkin IY, Woese C, Wiegel J (1994) Isolation and characterization of *Desulfitobacterium*  
27 *dehalogenans* gen. nov., sp. nov., an anaerobic bacterium which reductively dechlorinates  
28 chlorophenolic compounds. Int J Syst Bacteriol 44:612-619
- 29 56. Yoshida N, Asahi K, Sakakibara Y, Miyake K, Katayama A (2007) Isolation and quantitative  
30 detection of tetrachloroethene (PCE)-dechlorinating bacteria in unsaturated subsurface soils  
31 contaminated with chloroethenes. J Biosci Bioeng 104:91-97

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**Figure legends**

Fig. 1. Phylogenetic tree based on a comparison of 16S rRNA gene sequences. The phylogenetic tree was generated using neighbor-joining methods. Bootstrap values are shown from the bootstrap analysis of 100 replications. Scale bar represents an evolutionary distance (Knuc) of 0.01.

Fig. 2. The time courses of the ratios of residual to initial PCE (A) and *cis*-DCE (B) concentrations, and cell density (OD<sub>590</sub>). Concentrations used for PCE degradation and growth curve: closed diamonds, 0.06 mM; closed triangles, 0.12 mM; closed circles, 0.3 mM; closed squares, 0.6 mM. Concentrations used for *cis*-DCE degradation and growth curve: closed diamonds, 0.1 mM; closed triangles, 0.21 mM; closed circles, 0.52 mM; closed squares, 0.98 mM. Data represent means of triplicate experiments, and error bars indicate 95% confidence intervals.

Fig. 3. Effects of electron donors on PCE and *cis*-DCE degradation by cell-free extracts of strains HK-1 and HK-3. Initial protein concentration of crude enzyme was 0.79 mg ml<sup>-1</sup>. Results are means of triplicate independent experiments. Control experiment was conducted with enzyme and without electron donors. Data points are means of triplicate observations and error bars represent ±SD.

Fig. 4. Release of chloride ions during degradation of PCE (A) and *cis*-DCE (B) by cell-free extracts of strains HK-1 and HK-3. A) solid circles (PCE with enzyme); solid squares (PCE without enzyme); solid triangles (chloride ion). B) solid circles (*cis*-DCE with enzyme); solid squares (*cis*-DCE without enzyme); solid triangles (chloride ion). Initial protein concentration of crude enzyme was 1.2 mg ml<sup>-1</sup>. Data points are means of triplicate observations and error bars represent ±SD.

Fig. 5. Inactivation of PCE dehalogenase by propyl iodide and reactivation by light. The enzyme was reduced by the addition of 2 mM titanium (III) citrate (TC; arrow) prior to inactivation with 0.5 mM propyl iodide (PI). At 40 min, the assay mixture was exposed to light (hν). Relative activity was calculated compared with a control (without PI). One unit (U) of enzyme activity was defined as nmol of PCE degraded per hour under assay conditions. Results were normalized with respect to a maximal activity (100%) of 30 U. Initial protein concentration of crude enzyme was 1.2 mg ml<sup>-1</sup>. Results are means of duplicate experiments.

Figure 1

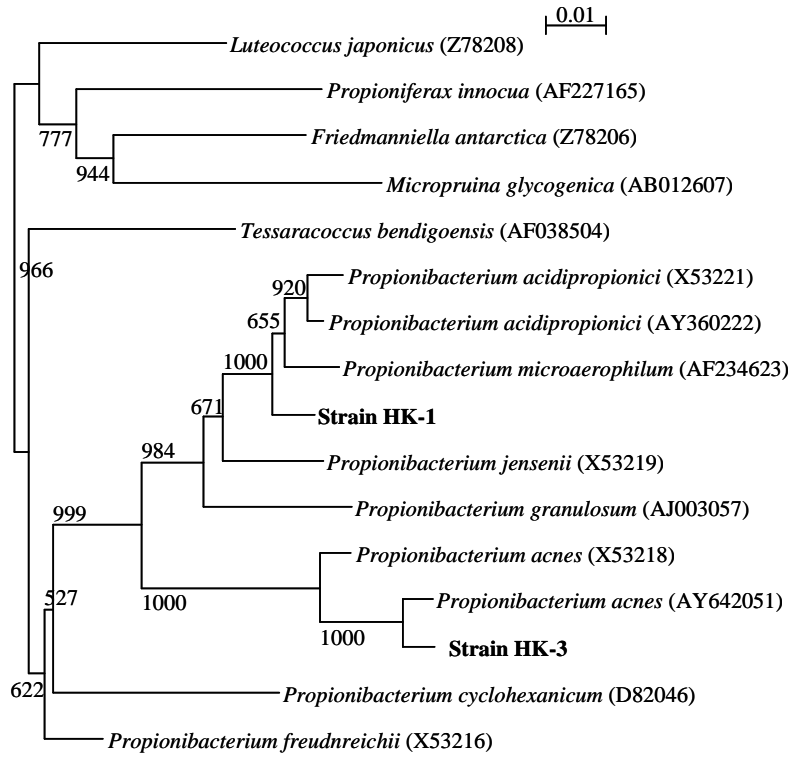


Figure 2.

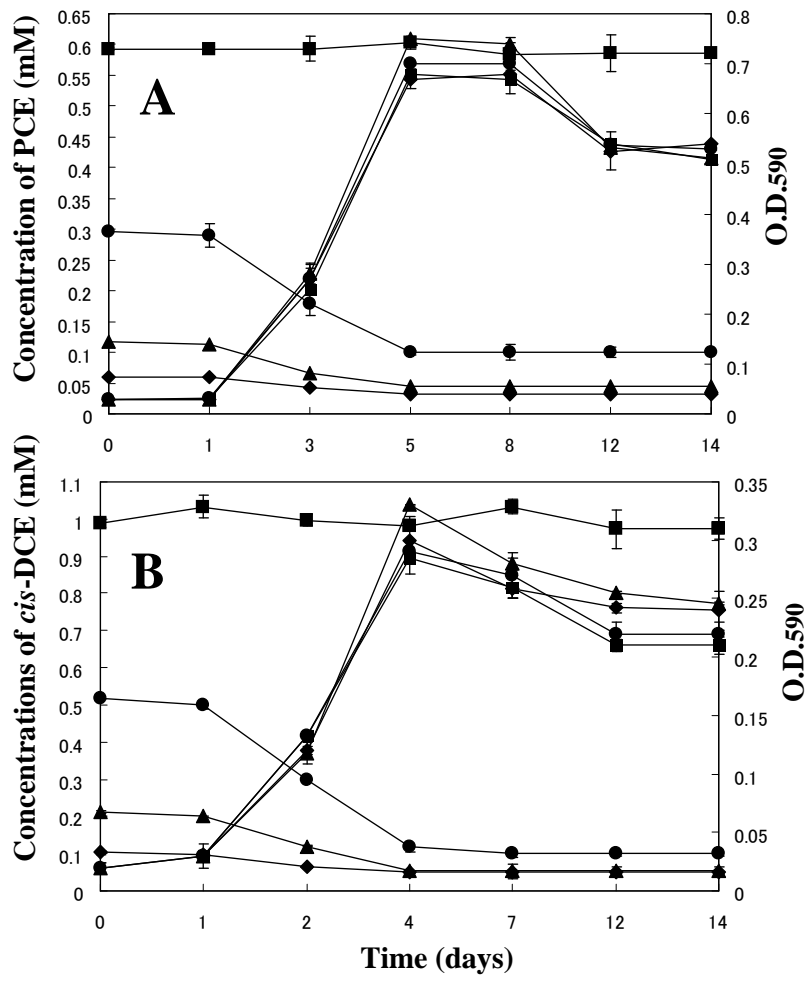


Figure 3.

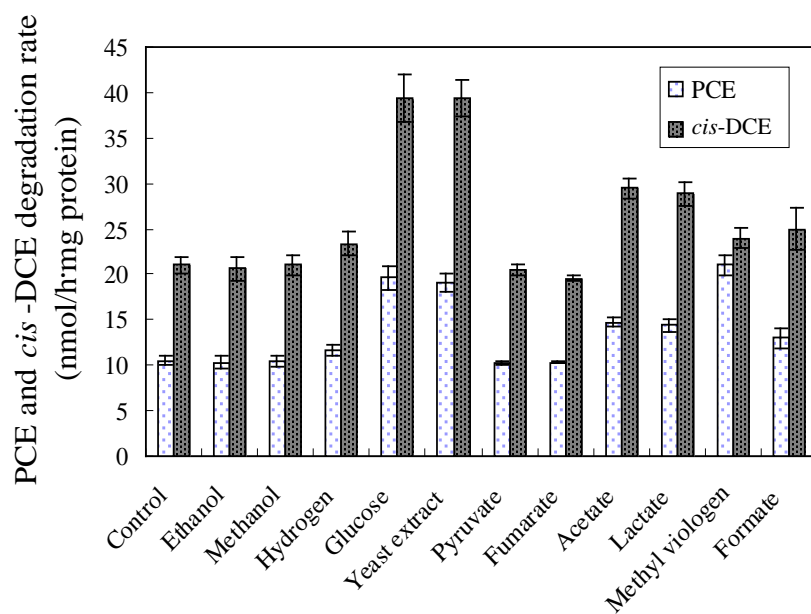


Figure 4.

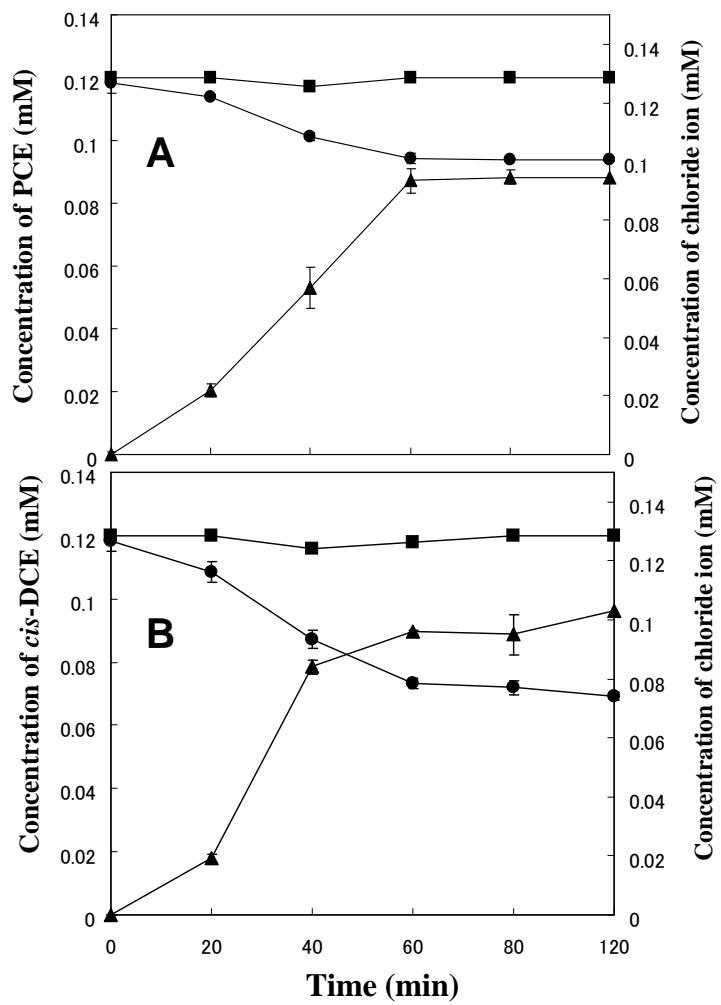


Figure 5.

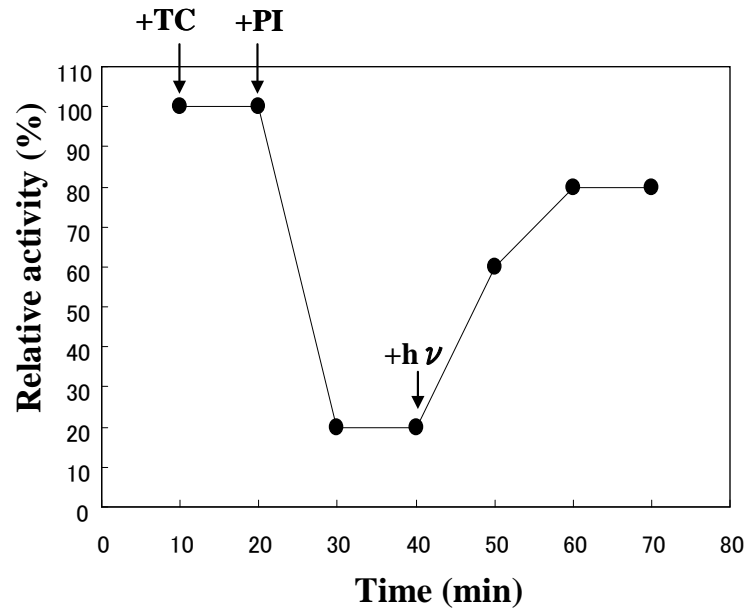
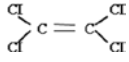
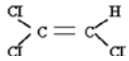
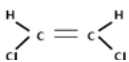
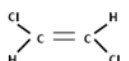
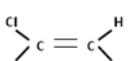
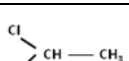
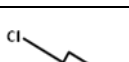

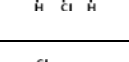
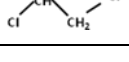


Table 1. Physiological characteristics of strains HK-1 and HK-3

Gram stain	Positive
Indole production	Negative
Protease hydrolysis	Negative
$\beta$ -glucosidase hydrolysis	Positive
Urease	Negative
Catalase	Positive
Oxidase	Positive
Utilization of carbon sources (Positive):	glucose, maltose, lactose, saccharose, L-arabinose, glycerol, D-cellobinose, D-mannose, D-melezitose, D-raffinose, D-sorbitol, D-rhamnose, D-trehalose, salicin, D-xylose, D-mannitol



Table 2. Biodegradation of chlorinated aliphatics by cell extracts<sup>a</sup>.

Halogenated aliphatic compound	Structure	Strain HK-1		Strain HK-3	
		Residual substrate( $\mu$ M)	Degradation (%) <sup>b</sup>	Residual substrate( $\mu$ M)	Degradation (%)
Tetrachloroethylene		96 $\pm$ 5.43	20 $\pm$ 1.13	120	ND <sup>c</sup>
Trichloroethylene		100.8 $\pm$ 7.46	16 $\pm$ 1.24	120	ND
<i>cis</i> -1,2-Dichloroethylene		90 $\pm$ 8.69	25 $\pm$ 2.26	78 $\pm$ 5.04	35 $\pm$ 2.26
<i>trans</i> -1,2-Dichloroethylene		100.8 $\pm$ 6.79	16 $\pm$ 1.13	98.4 $\pm$ 6.18	18 $\pm$ 1.13
1,1-Dichloroethylene		92.4 $\pm$ 5.66	23 $\pm$ 1.36	90 $\pm$ 12.22	25 $\pm$ 3.39
1,1-Dichloroethane		92.4 $\pm$ 5.10	23 $\pm$ 1.22	120	ND
1,2-Dichloroethane		92.4 $\pm$ 6.14	23 $\pm$ 1.13	120	ND
1,2-Dichloropropane		108 $\pm$ 10.86	10. $\pm$ 1.13	120	ND
1,1,2-Trichloroethane		90 $\pm$ 9.56	25 $\pm$ 2.49	120	ND
Vinyl chloride		84 $\pm$ 7.24	30 $\pm$ 2.26	72 $\pm$ 6.11	40 $\pm$ 3.39

<sup>a</sup>Biodegradation experiments were carried out in Tris-HCl (50 mM) with DTT (2 mM) and glucose (5 mM) as the electron donor at 30°C. Concentrations of remaining compounds were measured after 1 h. Initial concentration of protein was 1.2 mg ml<sup>-1</sup>. <sup>b</sup>Percent degradation compared with controls containing no enzyme. Values are means of triplicate experiments  $\pm$  SD. <sup>c</sup>ND: not degraded.